IBC Meeting Minutes

Chair- Ken Bondioli
Research Safety- Abigail Fish
Thursday, October 9, 2025
1:30 pm via Zoom

Institutions *Louisiana State University Agricultural and Mechanical College (A&M)*
Louisiana State University Ag Center

IBC Members			
	Ken Bondioli	LSU Ag Center	Chair, Animal Expert
	Abigail Fish	LSU A&M	BSO, Administrator, Voting Contact
	Sarah Keeton	LSU A&M	BSO
	Sue Hagius	LSU Ag Center	Animal Expert, Lab Rep
	Michael Hooks	LSU A&M	Member
	Jong Ham	LSU Ag Center	Plant Expert
	Christy White	Pennington Biomedical Research Center	Non-Voting Member
	Jeff Davis	LSU Ag Center	Plant and Insect Expert
	Niranjan Baisakh	LSU Ag Center	Plant Expert
	William Doerrler	LSU A&M	Member
	Ramanuj Lahiri	National Hansen's Disease Program	Member
	Rebecca Christofferson	LSU A&M	Member
	Michelle Dennis	Our Lady of the Lake Hospital	Local Non-Affiliated Member
	Brent Stanfield	LSU A&M	Member
	Ryoichi Teruyama	LSU A&M	Member

Members Present:	Ken Bondioli, Sue Hagius, Dipendra Shahi (proxy for Niranjan Baisakh), Abigail Fish, William Doerrler, Taylor Santaloci (proxy for Michael Hooks; left at 2:39 pm), Jong Ham, Ryoichi Teruyama, Samantha Clark (proxy for Rebecca Christofferson), and Sarah Keeton	
Members Absent:	Christy White, Michelle Dennis, Jeff Davis, Ramanuj Lahiri, and Brent Stanfield.	
Others Present:	Kristen Walker	Post-Doctoral Research, Museum of Natural Sciences
	Gregory Thom	Assistant Professor, Department of Biological Sciences Curator, Genetic Resources at the Museum of Natural Sciences
	Samithamby Jeyaseelan	Professor, Department of Pathobiological Sciences
	Brian Irving	Professor, School of Kinesiology
	Shafiqul Chowdhury	Professor, Department of Pathobiological Sciences
	Mandi Lopez	Professor, Department of Veterinary Clinical Sciences Director, Lab for Equine and Comparative Orthopedic Research
	Constantine Simintiras	Assistant Professor, School of Animal Sciences

Call to Order: 1:34 pm

Approval of Minutes from: ***Redacted** minutes from Thursday, June 12, 2025

Motion Made by:	Sue Hagius
Seconded by:	William Doerrler
Abstaining:	None

***Redacted** minutes from Thursday, July 17, 2025

Motion Made by:	Sue Hagius
Seconded by:	William Doerrler
Abstaining	None

The minutes from the September meeting are not complete. The committee is still pending revisions from Dr. Anne Grove (IBC 25054) and Dr. Fangneng Huang (IBC 25055). These individuals have until October 17, 2025, to submit revisions. If revisions are not received by then, conditional approval will be revoked.

Business and Call for New Business

The IBC Charter and Policies were updated, and committee members received a copy for review. Revisions and recommendations were received, and the document has been updated accordingly. A motion was made to accept the 2025 IBC Charter and Policies pending revisions.

Motion Made by: Sue Hagius

Seconded by: Sarah Keeton

The 2025 IBC Charter and Policies were approved pending revisions by the majority.

A new community member has been identified, and we are waiting for an official appointment by ORED. This individual's official appointment begins November 1, 2025.

The committee received a follow-up update regarding the previously reported suspension of funding for one LSU research project. Coordination between the Office of Research & Economic Development (ORED), LSU Legal Counsel, the research team, and biosafety personnel is ongoing to ensure the safe and compliant cessation of all related activities. The process remains in progress, and additional information will be communicated to the committee as it becomes available.

New IBC Registrations and Amendments for Review

Reg. #	PI Name	Affiliation of PI	Date Received	Title of Project	Reviewer 1	Reviewer 2
25047 (On Hold)	Robb Brumfield	Biological Sciences	9/24/2025	LSU Museum of Natural Sciences Research Collections	Sarah Keeton	William Doerrler

Project Overview:

The LSU Museum of Natural Science (LSUMNS) is dedicated to collecting, preserving, and studying specimens that help us understand the diversity of life on Earth, both past and present. Through active research, world-class collections, and robust educational programs, the museum plays a key role in advancing scientific knowledge and sharing it with students, researchers, and the public in Louisiana and beyond. To support its mission, the museum acquires vertebrate specimens, including birds and mammals, from around the world. Some of these collections originate in regions where diseases like Exotic Newcastle Disease are present, which requires special import permits regulated by the USDA. In order to legally and safely import these specimens, the museum must maintain Biosafety Level 2 (BSL-2) status. This project summary pertains to maintaining that status, which allows LSUMNS to continue its nationally and internationally recognized research and educational work in biodiversity and conservation.

Risk Assessment and Discussion:

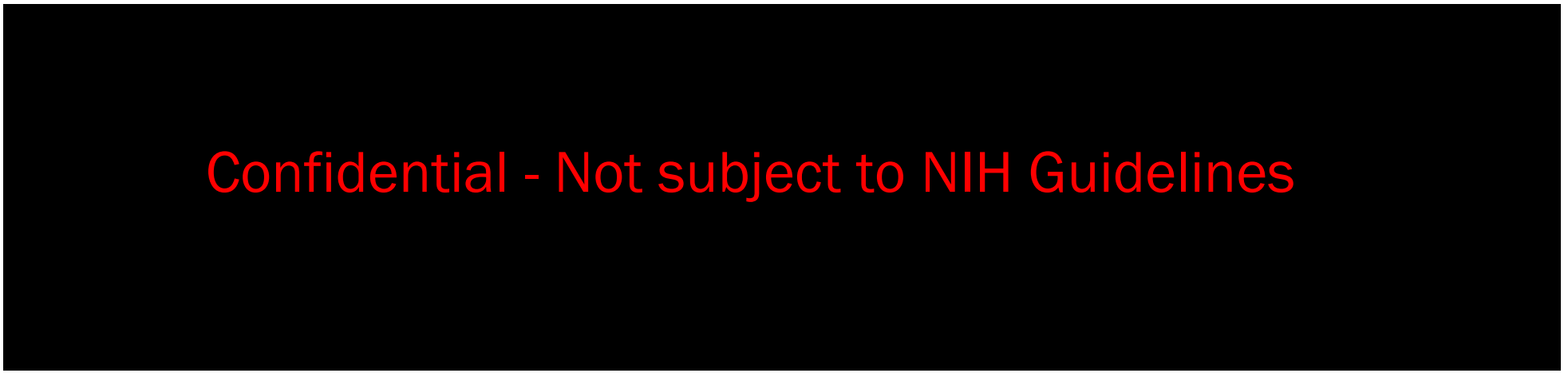
This project presents low overall risk but requires enhanced containment practices due to the nature and origin of the biological materials. The LSU Museum of Natural Science (LSUMNS) maintains vertebrate research collections that include birds and mammals obtained from both domestic and international sources. Some specimens may originate from areas where certain animal diseases—such as Exotic Newcastle Disease—are known to occur. While these specimens are preserved and not infectious under normal conditions, federal regulations require specific containment and handling procedures to ensure biosafety and compliance with USDA import permit requirements.

The primary risks relate to the potential presence of residual pathogens in animal tissues and the safe handling of these materials during collection, transport, and processing. These risks are effectively mitigated through LSU's **Biosafety Level 2 (BSL-2)** containment facilities, adherence to approved USDA permit conditions, and implementation of LSU's established biosafety policies and procedures. Personnel receive appropriate training, use personal protective equipment, and follow standard operating procedures for disinfection and waste disposal.

With these controls in place, the project poses minimal risk to researchers, the university community, and the public. The BSL-2 designation is a precautionary regulatory requirement that ensures continued safe and compliant operation of the museum's research and educational activities.

NIH Guidelines:	Not Applicable.
Biosafety Level:	BSL-2
Training Requirements:	All personnel, including the PI, involved in this project must complete BSL-2 training in accordance with LSU's Environmental Health and Safety (EHS) and Institutional Biosafety Committee (IBC) requirements. All training must be completed before beginning work and refreshed as required by LSU policies and SOPs.
IBC Vote:	**Approved at BSL-2 pending receipt of modifications**
Motion made by:	Sarah Keeton
Seconded by:	William Doerrler
Abstaining:	None
Conflicts of Interest:	None

Requested Modifications:



Reg. #	PI Name	Affiliation of PI	Date Received	Title of Project	Reviewer 1	Reviewer 2
25057 (Renewal)	Samithamby Jeyaseelan	Pathobiological Sciences	9/16/2025	Innate Defense Against Bacterial Pneumonia and Polymicrobial Sepsis Using Lentiviral Vectors	Michelle Dennis	Ryoichi Teruyama

Project Overview: The project was unclear due to limited information within the registration. The registration was returned to the PI for additional edits before full committee review.

Risk Assessment and Discussion: The IBC did not feel that this application contained enough information to complete a risk assessment; therefore, the registration was put "ON HOLD" until additional information is received.

NIH Guidelines: To be determined
Biosafety Level: To be determined
Training Requirements: To be determined once more information is provided.

IBC Vote: **No motion was made. The protocol was placed "ON HOLD".**

Motion made by:	Not applicable
Seconded by:	Not applicable
Abstaining:	Not applicable
Conflicts of Interest:	Not applicable

Requested Modifications:

- Section A. Project Information.
  - Buildings. Please update this section to reflect only the names of the buildings where work occurs.
  - Room numbers. Please list all room numbers, including DLAM room numbers, here.
  - Personnel. Training. Please have all lab personnel, including the PI, complete the EHS-required online BSL2 safety training and list the courses under specific training.
- Section B. Project Description.
  - Project Goals. Please spell out NLRP10 and NRF2.

    - Procedures and Methods. Please indicate what work takes place in each lab and what work is performed inside a BSC. Please describe recombinant DNA work. Briefly describe constructs and the viral vectors used. Please include the generation of the lentiviral vector. Please remove mention of the life sciences building if you no longer use this vivarium. Please describe animal work and state that transgenic mice are being used. Please be sure to include information on how animals are inoculated and tissue and sample collection. Please briefly describe procedures associated with the animal work. Importantly, please include information on how potentially infectious material is inactivated for each procedure. Please indicate how BAL is processed and briefly describe the cytokine analysis. Please be sure to include inactivation information for this procedure as well. Please describe bacterial culture techniques and indicate where this work is performed. Please elaborate on the procedure for flow cytometry. Be sure to include information on how the material is inactivated.
- Section C. Risk Evaluation.
    - Biosafety. Please indicate what PPE is worn when working in the lab and when handling animals. Please indicate what type of mask is being worn and when it is required. Please describe the use of BSCs. Please list other appropriate BSL2 practices for this work. If you have SOPs related to biosafety, the committee recommends that they be attached, and a summary of the procedures listed here.
    - Biosecurity. Please describe solid and liquid waste handling procedures. Please also indicate how animal carcasses are disposed of. Please describe building and lab security and inventory management.
- Section D. Project Units.
    - IACUC. Please update the IACUC protocol information, including the protocol number
- Section F. Recombinant DNA.
    - NIH Guidelines. Please update to Section III-D-4-b.
    - Source of DNA. Please include the species of coding sequences.
    - Gene products effects. Please clarify what “other issue” refers to.
    - Viral Components. Please list the vector, not the catalog number.
- Section G. Transgenic Animals.
    - Please list the strain/isolate for the transgenic animals used.
    - Please specify the nature of the transgenic rodents used under item 5.
    - Animal housing. Please provide room numbers if known and describe the containment measures for ABSL2.
    - Please confirm animal disposal information.
- Section J and K. Human and Animal Pathogens.
    - Please confirm you do not concentrate the pathogen. Centrifugation after culture is concentration.
- Section N. Safety.
    - Biosafety Cabinets. Please update certification information. If you have more than one BSC, please list them all here.
    - Personal Protective Equipment. If you plan to use masks, please check the appropriate type here. If not listed, please check other and indicate what type of mask is worn.
    - Other Safety Equipment. Please check Signs/Labels.
- Section O. Medical Surveillance.
    - Bloodborne Pathogens Training. Please change “no” to “yes” and ensure that all staff, including the PI, are up to date on BBP training. The use of lentiviral vectors required this training to be completed per the OSHA BBP standard.

Reg. #	PI Name	Affiliation of PI	Date Received	Title of Project	Reviewer 1	Reviewer 2
25058	Mark Mitchell	Veterinary Clinical Sciences	9/16/2025	Uncovering Health Issues and Conservation Needs for Louisiana's Critically Imperiled Gopher Tortoise (*Gopherus polyphemus*)	Sue Hagius	Jong Ham

Project Overview:

This research project seeks to better understand the health and disease ecology of the gopher tortoise, a keystone species native to the southeastern United States. Gopher tortoises play a vital role in their ecosystems, and understanding the factors that affect their health is essential for conservation and management.

To support these efforts, researchers will use diagnostic and molecular analysis techniques to examine the presence of several pathogens—including ranavirus, herpesvirus, and *Mycoplasma*—that may impact tortoise populations. The study will also analyze the normal bacterial and fungal communities (microbiomes) within the digestive system and identify common intestinal parasites. Together, these data will provide a more complete picture of the tortoise's overall health and help inform strategies to protect this important species and its habitat.

Risk Assessment and Discussion:

This project presents a low overall risk to personnel and the public. Researchers will conduct diagnostic and molecular analyses on gopher tortoise samples to detect *Mycoplasma*, ranavirus, herpesvirus, and other microorganisms. While these wildlife-associated agents are not significant human pathogens, reptiles—including tortoises—are known natural carriers of *Salmonella*, which can cause illness in humans if proper hygiene is not followed.

Risks are limited to standard laboratory and field activities, such as handling animal specimens, potential aerosol generation, and waste disposal. These are effectively managed through **Biosafety Level 2 (BSL-2) practices**, including use of biological safety cabinets, gloves and other PPE, disinfection of surfaces and tools, and proper biohazard waste disposal. Field biosecurity measures—such as changing gloves between animals and disinfecting equipment—further reduce the risk of cross-contamination or environmental spread.

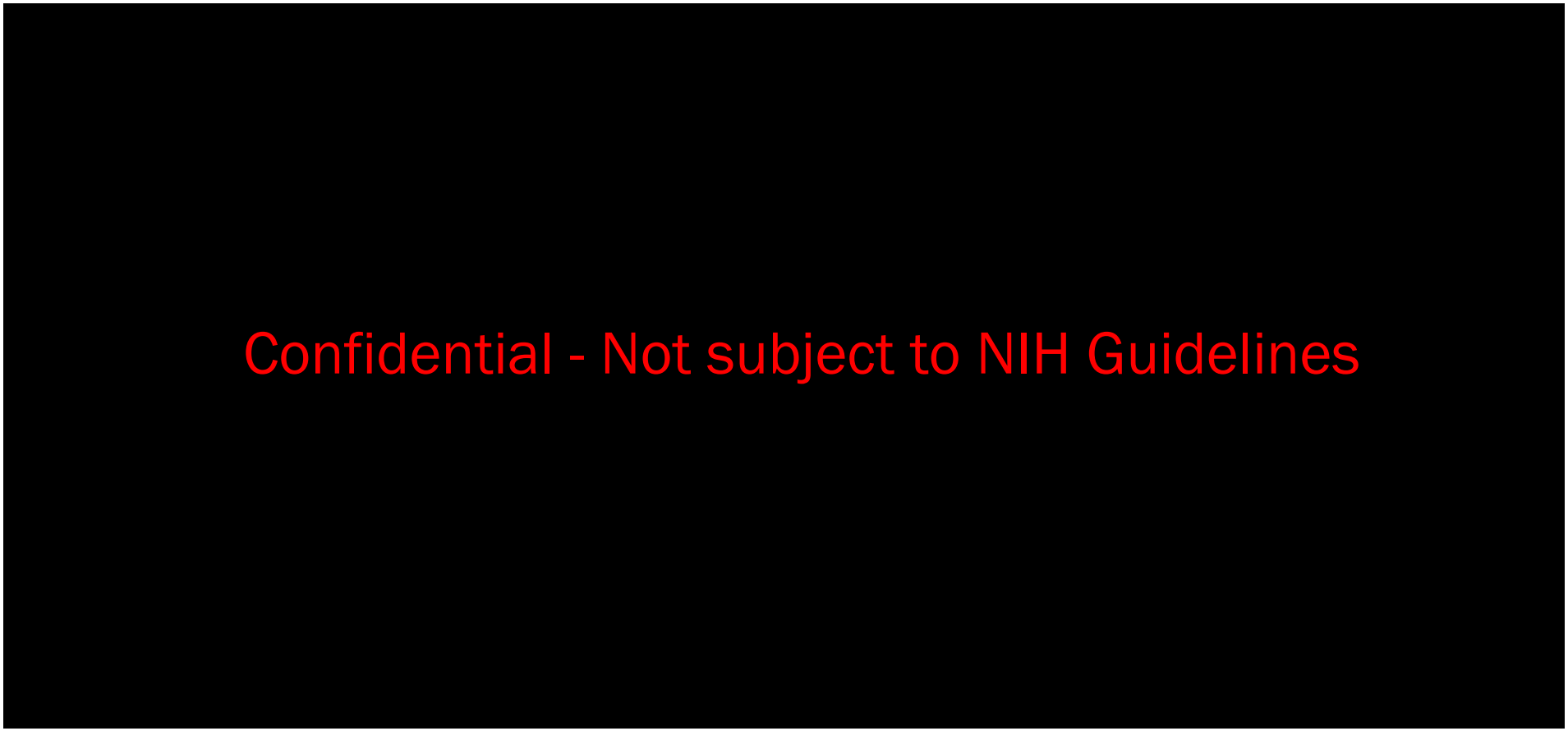
With these precautions in place, the project is considered low risk under standard biosafety guidelines, with no unusual safety, environmental, or security concerns anticipated.

NIH Guidelines: Not Applicable

Biosafety Level: BSL-2

Training Requirements:	All personnel, including the PI, involved in this project must complete BSL-2 training in accordance with LSU's Environmental Health and Safety (EHS) and Institutional Biosafety Committee (IBC) requirements. All training must be completed before beginning work and refreshed as required by LSU policies and SOPs.
IBC Vote:	**Approved at BSL-2 pending receipt of modifications and a lab inspection**
Motion made by:	Sue Hagius
Seconded by:	Jong Ham
Abstaining:	None
Conflicts of Interest:	None

Requested Modifications:



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Reg. #	PI Name	Affiliation of PI	Date Received	Title of Project	Reviewer 1	Reviewer 2
25059	William Doerrler	Biological Sciences	9/17/2025	SynrGNet2-ABX: Closing the Mutant Selection Window in Precision Antibiotic Combinations with Graph Neural Networks	Jong Ham	Ryoichi Teruyama

Project Overview:

This research project seeks to develop new computer-based tools to identify antibiotic combinations that can slow or prevent the development of bacterial resistance. Using a platform called SynerGNet-ABX, the research team will integrate advanced machine learning with laboratory testing to predict which antibiotic pairs work best together—both to kill bacteria effectively and to minimize the chance of resistant strains emerging.

To validate the computer predictions, the researchers will test various bacterial strains in the lab. They will determine the minimum inhibitory concentration (MIC) for different antibiotics and evaluate how pairs of drugs interact using checkerboard assays to measure synergy. Together, these approaches aim to accelerate the discovery of smarter, more sustainable treatment strategies that extend the usefulness of existing antibiotics.

Risk Assessment and Discussion:

This project presents a moderate biosafety risk consistent with work involving opportunistic or antibiotic-resistant bacteria. The research involves *Pseudomonas aeruginosa*, *Acinetobacter baumannii*, *Staphylococcus aureus*, *Klebsiella pneumoniae*, and *Escherichia coli*. Some of these strains may carry multidrug resistance or virulence traits. While they will be handled in small quantities, these organisms can cause infection in humans if accidentally introduced through cuts, mucous membranes, or aerosols.

All culture manipulations will be conducted under **Biosafety Level 2 (BSL-2)** containment using a certified biological safety cabinet. Personnel will wear appropriate PPE (lab coat, gloves, and eye protection) and follow LSU's biosafety and chemical hygiene protocols for antibiotic use, spill response, and waste disposal. Surfaces and materials will be decontaminated with EPA-approved disinfectants effective against Gram-negative and Gram-positive bacteria.

With these containment measures, the project is considered low to moderate risk under standard biosafety guidelines, and no environmental or security risks are anticipated.

NIH Guidelines:	Not Applicable
Biosafety Level:	BSL-2
Training Requirements:	All personnel, including the PI, involved in this project must complete BSL-2 training in accordance with LSU's Environmental Health and Safety (EHS) and Institutional Biosafety Committee (IBC) requirements. All training must be completed before beginning work and refreshed as required by LSU policies and SOPs.
IBC Vote:	**Approved at BSL-2 pending receipt of modifications**
Motion made by:	Jong Ham
Seconded by:	Ryoichi Teruyama
Abstaining:	William Doerrler
Conflicts of Interest:	None

Requested Modifications:

Reg. #	PI Name	Affiliation of PI	Date Received	Title of Project	Reviewer 1	Reviewer 2
25060	Constantine Simintiras	Animal Sciences	9/21/2025	Recombinant Mammalian Female Reproductive Tract Cell Culture	William Doerrler	Sarah Keeton

Project Overview:	This project seeks to better understand how genes regulate the normal function of the female reproductive tract, with a focus on the molecular pathways that control endometrial activity. The PI will accomplish this goal by culturing and performing targeted gene editing in mammalian reproductive tract cells—specifically endometrial and oviductal epithelial and stromal cells—and by maintaining previously generated recombinant mouse cell lines for comparison. The research will include culturing primary human and bovine reproductive tract cells and introducing recombinant DNA constructs using established gene-editing techniques such as lipofectamine-based transfection, electroporation, and lentiviral transduction. These approaches will allow the research team to investigate how specific genes influence cell behavior, communication, and tissue function. No infectious agents or biological toxins will be used. The findings are expected to advance understanding of reproductive biology and contribute to broader knowledge relevant to fertility, developmental health, and women's reproductive science.
Risk Assessment and Discussion:	This project presents low to moderate biosafety risk, consistent with mammalian cell culture and recombinant DNA work involving human-derived cells. Primary human female reproductive tract (FRT) cells and bovine cells, along with recombinant murine lines, will be manipulated using lipofectamine-based transfection, electroporation, and lentiviral vectors. No infectious pathogens or biological toxins are used. Key hazards include potential exposure to human-derived materials (treated as potentially infectious per OSHA Bloodborne Pathogens principles), lentiviral vectors, and chemical transfection reagents. Risks are mitigated under **BSL-2** containment using certified biological safety cabinets for all open manipulations; standard PPE (lab coat, gloves, eye/face protection as needed); hand hygiene; EPA-registered disinfectants; and proper biohazard waste handling. Personnel will follow LSU biosafety and chemical hygiene SOPs, complete required training (including BBP-aligned training for work with human materials), and use spill/exposure response procedures. All recombinant DNA activities will comply with the NIH Guidelines. With these controls, the project is considered low overall risk under standard biosafety guidelines, with no unusual environmental or security concerns anticipated.
NIH Guidelines:	Section III-D-1-a.
Biosafety Level:	BSL-2
Training Requirements:	All personnel, including the PI, involved in this project must complete BSL-2 training in accordance with LSU's Environmental Health and Safety (EHS) and Institutional Biosafety Committee (IBC) requirements. All training must be completed before beginning work and refreshed as required by LSU policies and SOPs.
IBC Vote:	**Approved at BSL-2 pending receipt of modifications** Motion made by: William Doerrler

Seconded by:	Sarah Keeton
Abstaining:	None
Conflicts of Interest:	None

Requested Modifications:

- Section B. Project Description.
  - Procedures and Methods. Please add a statement at the beginning of this section that all work takes place in the listed location and indicate what work takes place inside the BSC. Please briefly describe PPE and disinfection protocols. Please indicate what genes you plan to edit. Please indicate what generation lentiviral vectors you plan to use. Please add the room number for the freezer. Please define "standard molecular analyses." Please describe the use of human uterine fluid, organoids, stromal cells, and biopsy tissue here. Please include a statement that these items are obtained by a collaborator and shipped to LSU.
- Section C. Risk Evaluation.
  - Containment Level. Please uncheck ABSL-2 and BL2-N.
  - Biosecurity. Please include a statement on secure transport. If no potentially hazardous material is transported between labs, please state that in this section.
- Section F. Recombinant DNA.
  - DNA Guidelines. Please remove Section III-D-1-b and Section III-D-3. Please add section III-D-1-a.
  - DNA/RNA insertions/deletions. Please change "no" to "yes".
  - Please describe "mammalian genes of interest".
  - Will inserted genes be expressed? Please change "no" to "yes". And describe the toxicity of genes that you plan to express under item 3.
- Section M. Human or Primate Blood, Bodily Fluids, or Tissues.
  - Types of manipulations. Please check accordingly.
    Containment, Disposal, and Destruction Measures. Destruction of stocks. Please be sure to autoclave this material before you discard it. Please clarify how solid waste is discarded.
- Section N. Safety.
  - Disinfection/Decontamination. Please uncheck 10% bleach for solid waste.
  - Biosafety Cabinet. Please update certification information.
  - Other Safety Equipment. Please check fire extinguisher and phone.

Reg. #	PI Name	Affiliation of PI	Date Received	Title of Project	Reviewer 1	Reviewer 2
25061 (Renewal)	Shafiqul Chowdhury	Pathobiological Sciences	9/22/2025	Novel Pseudorabies Virus (PRV) Vectored Subunit Vaccine Against African Swine Fever	Sue Hagius	Rebecca Christofferson

Project Overview:	This project aims to develop and evaluate new vaccine candidates designed to protect pigs against African swine fever virus (ASFV), a highly contagious and economically significant livestock disease. The research focuses on generating and characterizing three versions of a novel pseudorabies virus quadruple mutant (PRVqmv) vaccine vector engineered to express different combinations of ASFV proteins. The first version will express eight ASFV proteins that form the core antigen cocktail, the second will include two additional ASFV proteins to broaden immune coverage, and the third will feature modified ASFV proteins in which certain B-cell epitope regions are selectively removed to refine the immune response. These recombinant PRV-based vaccine constructs will be characterized and validated in vitro before being shipped to collaborators at Ohio State University for vaccination and challenge studies in pigs. This work contributes to national efforts to develop safe, effective, and next-generation ASFV vaccines that could improve disease preparedness and protect swine health and agricultural stability.
Risk Assessment and Discussion:	This project presents minimal safety and security risks. The research involves laboratory generation and characterization of recombinant pseudorabies virus (PRV) vaccine vectors that express non-infectious African swine fever virus (ASFV) protein fragments. The work is limited to molecular cloning and cell culture activities conducted in vitro; no live ASFV or animal studies will occur at LSU. The primary risks are standard **Biosafety Level 2 (BSL-2)** laboratory considerations, including handling of viral vectors, biological samples, and routine molecular reagents. These are well controlled through LSU's established biosafety and viral vector safety protocols, including use of biological safety cabinets, personal protective equipment (PPE), and proper waste management. With these practices in place, no environmental or security risks are anticipated, and the overall project is considered low risk under standard biosafety guidelines.
NIH Guidelines:	Section III-D-1-a
Biosafety Level:	BSL-2

Training Requirements: All personnel, including the PI, involved in this project must complete BSL-2 training in accordance with LSU's Environmental Health and Safety (EHS) and Institutional Biosafety Committee (IBC) requirements. All training must be completed before beginning work and refreshed as required by LSU policies and SOPs.

IBC Vote: **Approved at BSL-2 pending receipt of modifications**

Motion made by:	Sue Hagius
Seconded by:	Sarah Keeton
Abstaining:	None
Conflicts of Interest:	None

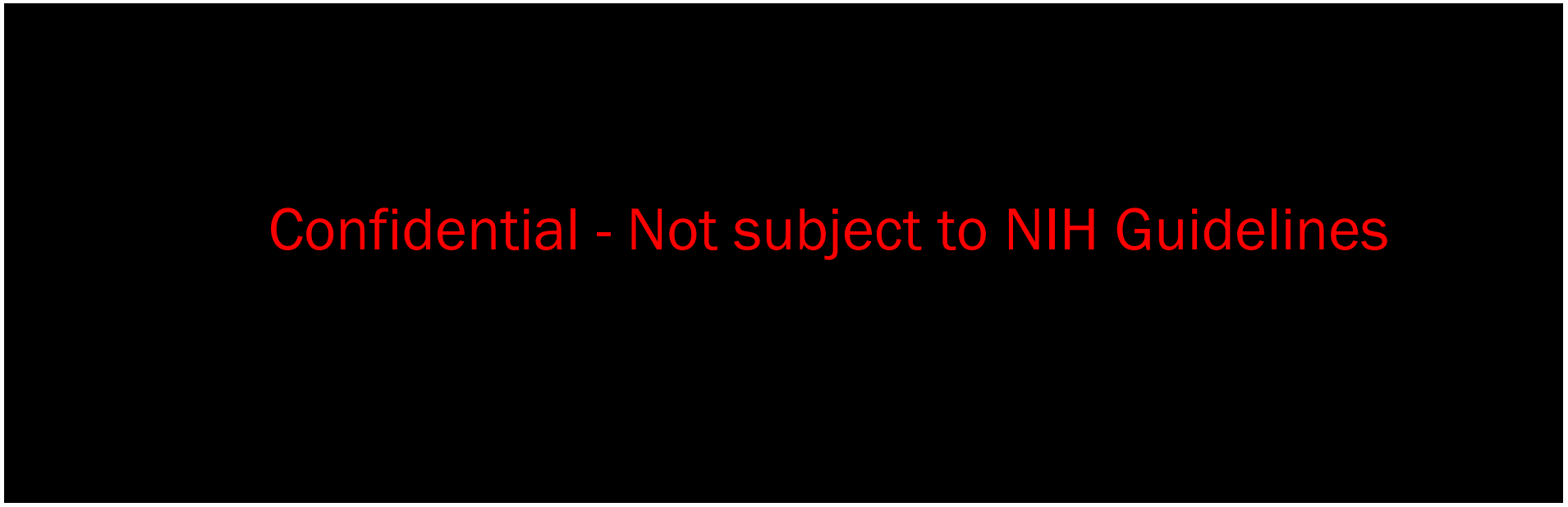
Requested Modifications:

- Section A. Project Information.
  - Personnel. Training. Please complete the EHS-required online BSL2 safety training and list the courses under specific training.
- Section B. Project Description.
  - Procedures and Methods. Please indicate what takes place in what lab and what work is performed inside a BSC. Please state that the wild-type virus will not be used. Please indicate when PRV and PRVtmv.CSFV.E2/Erns and PCV2B.Cap are used and for what. For all procedures, please indicate when potentially hazardous material is inactivated. Please briefly describe cell culture procedures and list relevant cell lines. Please describe viral propagation techniques and indicate what cells are used for viral packaging. Please briefly describe plaque assays, plaque purification, and immuno-blotting. Please describe recombinant DNA techniques, including transfections. Please include a statement at the beginning of this section that no animal work will be performed at LSU.
- Section F. Recombinant DNA.
  - DNA Guidelines. Please update to Section III-D-1-a.
  - Is the vector commercially available? Please list the company that supplies the plasmid.

Reg. #	PI Name	Affiliation of PI	Date Received	Title of Project	Reviewer 1	Reviewer 2
25062 (Renewal)	Brian Irving	Kinesiology	9/30/2025	The Effect of Altered Nitric Oxide Bioavailability on the Interaction with Obesity and Alzheimer's Disease	Abigail Fish	Michelle Dennis

Project Overview:	To be determined
Risk Assessment and Discussion:	To be determined
NIH Guidelines:	Not Applicable
Biosafety Level:	BSL-1
Training Requirements:	All personnel, including the PI, involved in this project must complete BSL-1 training in accordance with LSU's Environmental Health and Safety (EHS) and Institutional Biosafety Committee (IBC) requirements. All training must be completed before beginning work and refreshed as required by LSU policies and SOPs.
IBC Vote:	**Approved at BSL-1 pending receipt of modifications**
Motion made by:	Abigai Fish
Seconded by:	Sue Hagius
Abstaining:	None
Conflicts of Interest:	None

Requested Modifications:



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Reg. #	PI Name	Affiliation of PI	Date Received	Title of Project	Reviewer 1	Reviewer 2
25063	Mandi Lopez	Veterinary Clinical Sciences	9/30/2025	Effects of Cryopreservation, Immunophenotype, and Donor Sex on In-Vitro Behavior of Feline Adipose-Derived Stromal Cells	Sarah Keeton	Ryoichi Teruyama

Project Overview:

This research project seeks to better understand how certain factors influence the behavior of stem cells derived from cat fat tissue. These cells—called feline adipose-derived multipotent stromal cells—can grow into different types of tissues and are valuable for studying cell repair and regenerative medicine. By examining how freezing and thawing (cryopreservation), cell characteristics (immunophenotype), and sex affect their growth, flexibility, and gene activity, the researchers hope to learn how to preserve and use these cells more effectively in the future.

To do this, the PI will isolate and culture stem cells from feline adipose tissue, then compare how these cells grow and function under different conditions. The results may help improve storage and handling methods for veterinary or biomedical applications.

Risk Assessment and Discussion:

This project presents minimal safety and security risks. The research involves laboratory studies of feline adipose-derived multipotent stromal cells, which are non-infectious and commonly used in cell biology research. The work does not involve pathogenic agents, recombinant infectious materials, or hazardous genetic modifications. The procedures—such as isolating, culturing, freezing, and thawing cells—are standard tissue culture methods that do not increase risk to researchers or the public.

The primary risks are typical laboratory considerations, including safe handling of biological materials, use of chemical reagents for cell processing, and proper disposal of biohazardous waste. These risks are effectively managed under LSU's established biosafety and chemical safety programs. **Biosafety Level 1 (BSL-1)** practices are appropriate for this project, as it involves only low-risk materials and well-controlled laboratory techniques. No environmental or security risks are anticipated, and the overall project is considered low risk within standard biosafety guidelines.

NIH Guidelines: Not Applicable

Biosafety Level: BSL-1

Training Requirements: All personnel, including the PI, involved in this project must complete BSL-1 training in accordance with LSU's Environmental Health and Safety (EHS) and Institutional Biosafety Committee (IBC) requirements. All training must be completed before beginning work and refreshed as required by LSU policies and SOPs.

IBC Vote: **Approved at BSL-1 pending receipt of modifications**

Motion made by: Sarah Keeton

Seconded by: Ryoichi Teruyama

Abstaining: None

Conflicts of Interest: None

Requested Modifications:

[redacted]

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Reg. #	PI Name	Affiliation of PI	Date Received	Title of Project	Reviewer 1	Reviewer 2
25064	Hailey Parry	Kinesiology	10/1/2025	Investigating Mitochondria-Lipid Droplet Interaction in Skeletal Muscle	Ryoichi Teruyama	William Doerrler

Project Overview:	The project was unclear due to limited information within the registration. The registration was returned to the PI for additional edits before full committee review.
Risk Assessment and Discussion:	The IBC did not feel that this application contained enough information to complete a risk assessment; therefore, the registration was put "ON HOLD" until additional information is received.
NIH Guidelines:	To be determined
Biosafety Level:	To be determined
Training Requirements:	To be determined once more information is provided.

IBC Vote: **No motion was made. The protocol was placed "ON HOLD".**

Motion made by:	Not Applicable
Seconded by:	Not Applicable
Abstaining:	Not Applicable
Conflicts of Interest:	Not Applicable

Requested Modifications:

- Section A. Project Information.
  - Room Numbers. Please list room numbers for labs in Huey P. Long and Life Sciences.
  - Personnel. Training. Please have all lab personnel, including the PI, complete the EHS-required online BSL1 safety training and list the courses under specific training. Please add any staff or students working on this project. If no one is currently on the project, they may be added as amendments later.
- Section B. Project Description.
  - Project Goals. Please clarify project goals in laymen's terms.
  - Procedures and Methods. Please indicate what work takes place in what lab and what work is performed in a BSC. Please describe all procedures and include information on how potentially infectious material is inactivated for each procedure. Please describe recombinant DNA work. Please indicate what genetic material you plan to insert and into what plasmids. Please also briefly describe the gene function and the goal of the knock-in or knock-out. Please describe the animal work and indicate what type of mice you are using. Please include information on tissue collection and procedures with the samples post-necropsy. Please elaborate on imaging.
- Section C. Risk Evaluation.
  - Containment Level. Please check BL1-N.
  - Biosafety. Please list the PPE worn in the lab and when working with animals. Please indicate when a BSC is used. If no BSC is used, please describe the aerosol management plan. Please describe the training required for lab personnel.

    - Biosecurity. Please describe building and lab security for both buildings. Please add a statement indicating that only authorized individuals are permitted in the lab. Please detail how you handle solid and liquid waste. Please describe how animal carcasses are handled. Please include a statement on sharps. Please list the room number for the freezer. Please describe secure transport between labs and buildings.
- Section F. Recombinant DNA.
    - Source and nature of DNA/RNA inserts. Please identify the coding sequences by species and gene name.
    - Genes expressed. Please indicate if genes will be expressed in mice and E. coli.
    - Vectors. Please list vectors.
- Section N. Safety.
    - Sharps. Please check yes to scalpels if you will use them at necropsy. Please check yes to pipettes and pipette tips.
    - Disinfection/Decontamination. Please uncheck 10% bleach for solid waste and check the other appropriate box depending on the discard procedures.
    - Stock Cultures. Please confirm that stocks will not be kept after the study has been completed. Please list the room number for the freezer in HPL.
    - Personnel Protective Equipment. Please describe the use of PPE under Section C. Biosafety.

Upcoming Meetings: November 13, 2025 @1:30 pm via Zoom

Adjourned: 3:39 pm